

Biomarkers of Mild Cognitive Impairment and Alzheimer's Disease

Bor Luen Tang,¹*MSC, PhD*, Rajeev Kumar,²*MD, FRANZCP, PhD*

Abstract

Alzheimer's disease (AD) is currently diagnosed only via clinical assessments and confirmed by postmortem brain pathology. Biochemical and neuroimaging markers could facilitate diagnosis, predict AD progression from a pre-AD state of mild cognitive impairment (MCI), and be used to monitor efficacies of disease-modifying therapies. It is now clear that cerebrospinal fluid (CSF) levels of A β 40, A β 42, total tau and phosphorylated tau have diagnostic values in AD. Measurements of the above CSF markers in combination are useful in predicting the risk of progression from MCI to AD. Recent advances further support a notion that plasma A β levels, expressed as an A β 42/A β 40 ratio, could also be of value. New potential biomarkers are emerging, and CSF or plasma marker profiles may eventually become part of the clinician's toolkit for accurate AD diagnosis and management. These biomarkers, along with clinical assessment, neuropsychological testing and neuroimaging could achieve a much higher diagnostic accuracy for AD and related disorders in the future.

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Introduction

Alzheimer's disease (AD) is the most prevalent form of age-related dementia in the modern society.^{1,2} Other than symptomatic treatment with acetylcholinesterase inhibitors at its earlier stages,³ no disease-modifying strategies are currently known. AD diagnosis relies on clinical assessments and is often possible only when full dementia has set in. Reliable confirmation of AD diagnosis further depends on unhelpful postmortem brain pathology. Full AD dementia is usually preceded by a stage of cognitive decline, particularly with amnesia. This preclinical or prodromal AD state has been conceptualised as mild cognitive impairment (MCI), which has gained much attention as an ideal target for prevention and early intervention.

The term MCI is defined as a transitional clinical state between normal ageing and very mild AD.^{4,5} The diagnosis of MCI as a stable and valid concept in the community settings has been challenged by other researchers.⁶ Not all individuals with MCI progress to AD, and the ability to predict which ones would see such a progression greatly

facilitate the diagnosis of AD over normal ageing-associated cognitive impairment as well as other forms of dementia. It is clear that the discovery of clinically useful and robust biomarkers for AD and pre-AD are necessary for clinicians to accurately diagnose AD or predict conversion of a preclinical state of AD.

Alzheimer's Disease Pathology and Biomarker Candidates

Age-related, late-onset AD is largely idiopathic, and has 2 distinct pathological features – extracellular amyloid plaques and intracellular neurofibrillary tangles. The amyloid cascade hypothesis⁷ posits that the extracellular amyloid plaque, consisting of aggregated beta-amyloid (A β) peptide generated from proteolytic cleavages of the amyloid precursor protein (APP), damages brain regions and precipitates AD symptoms. An extension of this hypothesis is now necessary, as new findings indicate that both intracellular and extracellular soluble oligomeric forms of A β could result in synaptic malfunctions and the onset of AD.⁸⁻¹¹

¹ Department of Biochemistry

² Department of Psychological Medicine

Yong Loo Lin School of Medicine, National University of Singapore, Singapore

Address for Correspondence: Dr Tang Bor Luen, Department of Biochemistry, Yong Loo Lin School of Medicine, National University of Singapore, 8 Medical Drive, Singapore 117597.

Email: bchtbl@nus.edu.sg; pcmrk@nus.edu.sg

A β generation from APP occurs when the β -site APP-cleaving enzyme (BACE-1)¹² cleaves the ectodomain of APP to first generate a membrane-bound C-terminal fragment. Another subsequent cleavage by γ -secretase activity¹³ further generates peptides, mainly of 40 or 42 amino acids in length, termed A β 40 and A β 42. Both species could be found in amyloid plaques, but the latter is more directly neurotoxic and has a greater propensity to aggregate,¹⁴ while the former may actually be neuroprotective.¹⁵ Known genetic predispositions to early-onset AD include mutations in APP and the presenilins (part of the γ -secretase complex), and all of which increase A β generation or A β 42/A β 40 ratio.¹⁶ Other than APP and presenilin mutations, the ϵ 4 allele of apolipoprotein E (ApoE) also constitute a risk factor for late-onset AD.¹⁷

NFT, on the other hand, are intracellular filamentous aggregates of the microtubule binding protein tau. In AD and other tauopathies¹⁸ tau becomes hyperphosphorylated and accumulates into insoluble paired helical filaments, which aggregate into NFT.¹⁹ Like A β , NFT levels correlate well with neuron loss and cognitive impairment in AD patients. The trigger for tau hyperphosphorylation and aggregation in AD is yet unclear, although it would presumably involve an aberrant imbalance of tau phosphorylation and dephosphorylation, and which could be facilitated by A β .²⁰

An AD biomarker should ideally have the following features.^{21,22} It should detect a fundamental feature of AD neuropathology, with results that could be validated in neurologically confirmed cases. It should have a sensitivity of >85% and a specificity of >75%, and should be precise, reliable, inexpensive, convenient and with low risk to patients. Such idealised biomarkers may never be discovered. However, even those that partially fulfilled the above criteria would aid both predictive AD diagnoses from MCI presentations, as well as the monitoring of efficacies of disease-modifying therapies on trial. Recent advances have reaffirmed that both cerebrospinal fluid (CSF) A β and tau could serve as biomarkers for AD, and that plasma A β profiles, if appropriately measured, could also be promising. We highlight some of these findings in the paragraphs below, and discuss the possibility of applying these biomarker measurements to both cross-sectional and longitudinal cohort studies in Singapore.

Cerebrospinal Fluid A β and Tau as Biomarkers of AD and Incipient AD

The A β 40 and A β 42 peptides mentioned above are found in amyloid plaques and could form synapse-damaging oligomers. In the CSF, A β 40, A β 42 and other minor forms of peptides generated from APP (e.g., A β 37 and A β 38) could be detected and measured by immunochemical methods (such as ELISA) or liquid chromatography-mass

spectrometry. Likewise, tau (in its hyperphosphorylated forms), the principle protein in the neurofibrillary tangles, could be detected in CSF. A majority of findings have, however, indicated that CSF levels of A β 40 exhibited no significant differences, and have a large degree of overlapping, between AD patients and controls. On the other hand, A β 42 levels were generally noted to be decreased in AD patients, up to about half of that of controls.²³⁻²⁶ This parameter has both sensitivity and specificity in differentiating between AD patients and cognitively normal controls, but not between AD and other forms of dementia (e.g., vascular dementia, Creutzfeldt-Jakob disease and fronto-temporal dementia). This is likewise the case for total tau.²³⁻²⁶ More recent works have analysed specific phosphorylated forms of tau using phosphor-epitope-specific antibodies, and the results are also largely indicative of a high level of sensitivity and specificity.²⁷⁻²⁹

Encouragingly, recent studies have also indicated that both A β 42 and tau have significant predictive powers in cases of MCI progressing to AD. In particular, simultaneous measurement of CSF levels of A β peptides and tau, and expressing these levels as various ratios, help to increase the sensitivity and specificity of prediction.³⁰⁻⁴⁰ There are also studies with findings that indicate the ability of these markers to tell AD apart from other forms of dementia.⁴¹⁻⁴⁵ CSF A β and tau could also be promising antecedent biomarkers that could predict the future development of dementia in cognitively normal older adults.⁴⁵

Plasma A β and AD

CSF collection is invasive and unlikely to become a routine procedure in geriatric clinics. Finding peripheral biomarkers for AD is therefore of great interest. However, the levels of tau in the plasma are too low for any useful analysis. A β levels, while detectable, are also at least a magnitude lower. Earlier studies did not reveal significant diagnostic values for plasma A β peptides.^{46,47} Unlike changes in the CSF, reports of changes in A β levels in AD and pre-AD are rather inconsistent,⁴⁸⁻⁵⁰ and plasma levels do not necessarily reflect that in the brain.⁵¹ In spite of these difficulties, several recent reports have now increased the confidence that plasma A β may be of diagnostic value.

The Rotterdam Study is one of the largest ongoing prospective population-based cohort studies on the incidence and risk factors for age-related diseases, unique both in terms of its size and long-term follow-up. Van Oijen and colleagues found in this cohort an association between high A β 40 and low A β 42 levels and risk for AD dementia.⁵² Another study, which compared plasma A β 42 levels of 146 sporadic AD patients, 89 subjects with MCI and 89 age-matched controls found that a reduction in A β 42 is predictive for AD, and specifically, a transition from a normal state of cognition or MCI to AD.⁵³ A recent report

also indicated that while plasma A β 42 levels alone may not be good enough as a biomarker, it is increased in early AD and changes in its levels could indicate a transition from MCI to AD.⁵⁴ In another cohort with long-term follow-up, the plasma A β 42/A β 40 ratio was shown to be a useful biomarker for identifying cognitively normal elderly white subjects at risk for developing MCI or AD.⁵⁵

The above studies have changed the outlook of plasma A as an AD biomarker from an earlier perceived status of being “not very useful” to at least “moderately promising”. More studies are clearly warranted, as accurate and precise measurements of plasma A β is riddled with uncertainties and confounding factors. The non-specific binding capacity of A β 42 to proteins in the plasma is notorious. These bindings could mask detectable epitopes, and the degree of this masking could vary with metabolic conditions that differ from one subject to another. One other major uncertainty in the detection and measurements of plasma A β is the nature of the species measured by the antibodies used – whether they are monomeric, oligomeric or both.⁵⁶ Standardisation of studies using well-characterised antibodies with known A β species specificities would help. A promising possibility along this line of thought is that risks of MCI to AD could be further tiered by, for example, the ratio of oligomeric to monomeric A β .

Emerging Biomarkers for MCI and AD

Several other candidates for AD biomarkers have shown some initial promise but have not received as much attention as A β and tau. Lipid peroxidation is known to be a rather early event in AD,⁵⁷ and the measurement of CSF F2-isoprostane levels may, in combination with other parameters, serve to predict AD.⁵⁸ A rather accessible peripheral marker is platelet amyloid precursor protein (APP) isoform. An altered pattern of platelet APP isoform, manifested by a reduced ratio of the upper (130 kDa) to the lower (110 to 106 kDa) APP band, is associated with AD.⁵⁹ Interestingly, this platelet APP isoform ratio has also shown some documented predictive power in terms of MCI to AD progression.^{60,61}

Advances in the techniques of whole proteome analysis have naturally prompted the search of AD biomarkers using this particular approach.⁶² Comparative proteomics analysis with the aim of discovering proteins whose levels might be altered in diseased compared to control states, have been largely focused on CSF.⁶³⁻⁶⁶ The outcome of such analysis could be the identification of not 1 or 2, but a panel of proteins whose levels may be altered between normal controls, MCI subjects and cases of clinically diagnosed AD dementia.^{67,68} This sort of profiling analysis could increase sensitivity and specificity, particularly in discriminating AD from other forms of dementia.

Although plasma biomarkers are more clinically useful,

there are potentially confounding problems with biomarker mining. One prominent problem is that a small number of abundant proteins [such as albumin, α 2-macroglobulin (α 2-M) and immunoglobulins] could account for as much as 80% of the total protein in plasma. The large quantity of these proteins makes it difficult to identify low-abundance proteins in serum using traditional 2-dimensional electrophoresis. One way to go about this is to selectively deplete these abundant proteins, but there is a possibility that candidate biomarkers, already in low abundance, could be co-depleted. In view of the above difficulties, a recent report on novel possible plasma biomarkers discovered by a proteomics approach is of interest.⁶⁹ The authors reported that the levels of 2 proteins, complement factor H (CFH), and an abundant serum protein, α 2-M, are elevated in AD plasma. Preliminary analysis suggests that these elevations may correlate with disease severity. Both proteins have at least some remote relationship with AD pathology. α 2-M can be induced by inflammatory cytokines, is present in amyloid plaques, and competes with ApoE for the same receptor. CFH has also been previously shown to be present in plaques, and its encoding gene has been strongly associated with age-related macular degeneration, which shares some pathological features with AD. Although further studies will be necessary to fully validate the usefulness of α 2-M and CFH as AD or pre-AD biomarkers, the fact that these could be analysed in plasma makes them attractive from a clinical sampling point.

While there is a growing interest in studying biomarkers, there is still a scarcity of data in this area. It is also important to recognise that at this stage, biomarkers can only be used as a research tool and the data from small clinical populations on these still cannot be generalised to the general population. A way to move forward is to study existing biomarkers as well as to develop newer ones and test them in large representative samples of AD subjects. In order to be used widely as a diagnostic or prognostic marker, a candidate marker has to be studied both in clinical as well as epidemiological settings. Ideally, such studies should not only be collaborative in nature but should also include excellent clinical and neuropsychological assessments, cutting-edge neuroimaging and neurophysiological techniques, and genetic and genomic studies. Ultimately, instead of relying heavily on solitary markers alone, only a combined multimodal approach is going to be effective.

Studying MCI and AD in Singapore’s Ageing Population

Singapore is entering into an exciting time of neuroscience research. Policy makers and international advisory panels have rightly identified neuroscience research as an important strategic area of development, and have encouraged both clinical as well as basic scientists to work hand-in-hand to tackle age-related neurodegenerative diseases. Singapore

has an ageing population that would facilitate community-based, cohort studies. One such ongoing ageing cohort study is the Singapore Longitudinal Ageing Study (SLAS).^{70,71} There are also much interest and expertise in longitudinal monitoring of the cognitive impairment after stroke and vascular dementia,^{72,73} as well as preclinical AD.^{74,75}

There is clearly a growing interest among clinicians and basic scientists to tap on each other's expertise in the area of ageing neurobiology research. Such collaborations between geriatricians, neuroimaging specialists, neuropsychiatrists as well as molecular and cellular neurobiologists are being fostered. Streamlining research initiatives in a way that would maximise subject resources, data acquisition and multifaceted analyses should be of high priority. The prospect of seeing how the above CSF and plasma biomarkers correlate with the clinical findings, stratified ethnically, is an exciting one.

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REFERENCES

- Yaari R, Corey-Bloom J. Alzheimer's disease. *Semin Neurol* 2007;27:32-41.
- Schott JM, Kennedy J, Fox NC. New developments in mild cognitive impairment and Alzheimer's disease. *Curr Opin Neurol* 2006;19:552-8.
- Holzgrabe U, Kapkova P, Alptuzun V, Scheiber J, Kugelmann E. Targeting acetylcholinesterase to treat neurodegeneration. *Expert Opin Ther Targets* 2007;11:161-79.
- Petersen RC, Doody R, Kurz A, Mohs RC, Morris JC, Rabins PV, et al. Current concepts in mild cognitive impairment. *Arch Neuro* 2001;58:1985-1992.
- Peterson RC. Mild cognitive impairment: current research and clinical implications. *Semin Neurol* 2007;27:22-31.
- Ritchie K, Artero S, Touchon J. Classification criteria for mild cognitive impairment: a population-based validation study. *Neurology* 2001;56:37-42.
- Hardy J, Selkoe DJ. The amyloid hypothesis of Alzheimer's disease: progress and problems on the road to therapeutics. *Science* 2002;297:353-6. Erratum in: *Science* 2002;297:2209.
- Selkoe DJ. Alzheimer's disease is a synaptic failure. *Science* 2002;298:789-91.
- Wirhth O, Multhaup G, Bayer TA. A modified β -amyloid hypothesis: intraneuronal accumulation of the β -amyloid peptide—the first step of a fatal cascade. *J Neurochem* 2004;91:513-20.
- Cuello AC. Intracellular and extracellular A β , a tale of two neuropathologies. *Brain Pathol* 2005;15:66-71.
- Lacor PN, Buniel MC, Furlow PW, Clemente AS, Velasco PT, Wood M, et al. A β oligomer-induced aberrations in synapse composition, shape, and density provide a molecular basis for loss of connectivity in Alzheimer's disease. *J Neurosci* 2007;27:796-807.
- Dominguez DI, Hartmann D, De Strooper B. BACE1 and presenilin: two unusual aspartyl proteases involved in Alzheimer's disease. *Neurodegener Dis* 2004;1:168-74.
- Haass C. Take five-BACE and the gamma-secretase quartet conduct Alzheimer's amyloid beta-peptide generation. *EMBO J* 2004;23:483-8.
- Zhang Y, McLaughlin R, Goodyear C, LeBlanc A. Selective cytotoxicity of intracellular amyloid- β peptide 1-42 through p53 and Bax in cultured primary human neurons. *J Cell Biol* 2002;156:519-29.
- Zou K, Kim D, Kakio A, Byun K, Gong JS, Kim J, et al. Amyloid β -protein (A β)1-40 protects neurons from damage induced by A β 1-42 in culture and in rat brain. *J Neurochem* 2003;87:609-19.
- Tanzi RE, Bertram L. Twenty years of the Alzheimer's disease amyloid hypothesis: a genetic perspective. *Cell* 2005;120:545-55.
- Huang Y, Weisgraber KH, Mucke L, Mahley RW. Apolipoprotein E: diversity of cellular origins, structural and biophysical properties, and effects in Alzheimer's disease. *J Mol Neurosci* 2004;23:189-204.
- Iqbal K, Alonso Adel C, Chen S, Chohan MO, El-Akkad E, Gong CX, et al. Tau pathology in Alzheimer disease and other tauopathies. *Biochim Biophys Acta* 2005;1739:198-210.
- Mi K, Johnson GV. The role of tau phosphorylation in the pathogenesis of Alzheimer's disease. *Curr Alzheimer Res* 2006;3:449-63.
- Blurton-Jones M, Laferla FM. Pathways by which Abeta facilitates tau pathology. *Curr Alzheimer Res* 2006;3:437-48.
- The Ronald and Nancy Reagan Research Institute of the Alzheimer's Association and the National Institute on Aging Working Group. Consensus report of the Working Group on: "Molecular and Biochemical Markers of Alzheimer's Disease". *Neurobiol Aging* 1998;19:109-16. Erratum in: *Neurobiol Aging* 1998;19:285.
- Growdon JH. Biomarkers of Alzheimer disease. *Arch Neurol* 1999;56:281-3.
- Sjogren M, Andreasen N, Blennow K. Advances in the detection of Alzheimer's disease—use of cerebrospinal fluid biomarkers. *Clin Chim Acta* 2003;332:1-10.
- Blennow K, Hampel H. CSF markers for incipient Alzheimer's disease. *Lancet Neurol* 2003;2:605-13.
- Hampel H, Mitchell A, Blennow K, Frank RA, Brettschneider S, Weller L, et al. Core biological marker candidates of Alzheimer's disease – perspectives for diagnosis, prediction of outcome and reflection of biological activity. *J Neural Transm* 2004;111:247-72.
- Galasko D. Biomarkers for Alzheimer's disease – clinical needs and application. *J Alzheimers Dis* 2005;8:339-46.
- Itoh N, Arai H, Urakami K, Ishiguro K, Ohno H, Hampel H, et al. Large-scale, multicenter study of cerebrospinal fluid tau protein phosphorylated at serine 199 for the antemortem diagnosis of Alzheimer's disease. *Ann Neurol* 2001;50:150-6.
- Buerger K, Zinkowski R, Teipel SJ, Tapiola T, Arai H, Blennow K, et al. Differential diagnosis of Alzheimer disease with cerebrospinal fluid levels of tau protein phosphorylated at threonine 231. *Arch Neurol* 2002;59:1267-72.
- Hampel H, Buerger K, Zinkowski R, Teipel SJ, Goernitz A, Andreasen N, et al. Measurement of phosphorylated tau epitopes in the differential diagnosis of Alzheimer disease: a comparative cerebrospinal fluid study. *Arch Gen Psychiatry* 2004;61:95-102.
- Andreasen N, Minthon L, Vanmechelen E, Vanderstichele H, Davidsson P, Winblad B, et al. Cerebrospinal fluid tau and A β 42 as predictors of development of Alzheimer's disease in patients with mild cognitive impairment. *Neurosci Lett* 1999;273:5-8.
- Maruyama M, Arai H, Sugita M, Tanji H, Higuchi M, Okamura N, et al. Cerebrospinal fluid amyloid beta (1-42) levels in the mild cognitive impairment stage of Alzheimer's disease. *Exp Neurol* 2001;172:433-6.
- Hampel H, Teipel SJ, Fuchsberger T, Andreasen N, Wiltfang J, Otto M, et al. Value of CSF beta-amyloid 1-42 and tau as predictors of Alzheimer's disease in patients with mild cognitive impairment. *Mol Psychiatry* 2004;9:705-10.
- Formichi P, Battisti C, Radi E, Federico A. Cerebrospinal fluid tau, A β , and phosphorylated tau protein for the diagnosis of Alzheimer's disease. *J Cell Physiol* 2006;208:39-46.
- Herukka SK, Hallikainen M, Soininen H, Pirttila T. CSF A β 42 and tau or phosphorylated tau and prediction of progressive mild cognitive impairment. *Neurology* 2005;64:1294-7.
- De Leon MJ, DeSanti S, Zinkowski R, Mehta PD, Pratico D, Segal S, et al. Longitudinal CSF and MRI biomarkers improve the diagnosis of mild cognitive impairment. *Neurobiol Aging* 2006;27:394-401.

36. Hansson O, Zetterberg H, Buchhave P, Londos E, Blennow K, Minthon L. Association between CSF biomarkers and incipient Alzheimer's disease in patients with mild cognitive impairment: a follow-up study. *Lancet Neurol* 2006;5:228-34. Erratum in: *Lancet Neurol* 2006;5:293.
37. Herukka SK, Helisalml S, Hallikainen M, Tervo S, Soinen H, Pirttila T. CSF A β 42, Tau and phosphorylated Tau, APOE epsilon4 allele and MCI type in progressive MCI. *Neurobiol Aging* 2007;28:507-14.
38. Hansson O, Zetterberg H, Buchhave P, Andreasson U, Londos E, Minthon L, et al. Prediction of Alzheimer's disease using the CSF A β 42/A β 40 ratio in patients with mild cognitive impairment. *Dement Geriatr Cogn Disord* 2007;23:316-20.
39. Bouwman FH, Schoonenboom SN, van der Flier WM, van Elk EJ, Kok A, et al. CSF biomarkers and medial temporal lobe atrophy predict dementia in mild cognitive impairment. *Neurobiol Aging* 2007;28:1070-4.
40. Lewczuk P, Kornhuber J, Vanderstichele H, Vanmechelen E, Esselmann H, Bibl M, et al. Multiplexed quantification of dementia biomarkers in the CSF of patients with early dementias and MCI: A multicenter study. *Neurobiol Aging* 2008;29:812-8.
41. Maddalena A, Papassotiropoulos A, Muller-Tillmanns B, Jung HH, Hegi T, Nitsch RM, et al. Biochemical diagnosis of Alzheimer disease by measuring the cerebrospinal fluid ratio of phosphorylated tau protein to β -amyloid peptide 42. *Arch Neurol* 2003;60:1202-6.
42. Kapaki E, Paraskevas GP, Zalonis I, Zournas C. CSF tau protein and beta-amyloid (1-42) in Alzheimer's disease diagnosis: discrimination from normal ageing and other dementias in the Greek population. *Eur J Neurol* 2003;10:119-28.
43. Schoonenboom NS, Pijnenburg YA, Mulder C, Rosso SM, Van Elk EJ, Van Kamp GJ, et al. Amyloid β (1-42) and phosphorylated tau in CSF as markers for early-onset Alzheimer disease. *Neurology* 2004;62:1580-4.
44. Kapaki EN, Paraskevas GP, Tzerakis NG, Sfagos C, Seretis A, Kararizou E, et al. Cerebrospinal fluid tau, phospho-tau181 and β -amyloid 1-42 in idiopathic normal pressure hydrocephalus: a discrimination from Alzheimer's disease. *Eur J Neurol* 2007;14:168-73.
45. Fagan AM, Roe CM, Xiong C, Mintun MA, Morris JC, Holtzman DM. Cerebrospinal fluid tau/ β -amyloid 42 ratio as a prediction of cognitive decline in nondemented older adults. *Arch Neurol* 2007;64:343-9.
46. Mehta PD, Pirttila T, Mehta SP, Sersen EA, Aisen PS, Wisniewski HM. Plasma and cerebrospinal fluid levels of amyloid proteins 1-40 and 1-42 in Alzheimer disease. *Arch Neurol* 2000;57:100-5.
47. Mayeux R, Honig LS, Tang MX, Manly J, Stern Y, Schupf N, et al. Plasma A β 40 and A β 42 and Alzheimer's disease: relation to age, mortality, and risk. *Neurology* 2003;61:1185-90.
48. Fukumoto H, Tennis M, Locascio JJ, Hyman BT, Growdon JH, Irizarry MC. Age but not diagnosis is the main predictor of plasma amyloid beta-protein levels. *Arch Neurol* 2003;60:958-64.
49. Borroni B, Di Luca M, Padovani A. Predicting Alzheimer dementia in mild cognitive impairment patients. Are biomarkers useful? *Eur J Pharmacol* 2006;545:73-80.
50. Irizarry MC. Biomarkers of Alzheimer disease in plasma. *NeuroRx* 2004;1:226-34.
51. Freeman SH, Raju S, Hyman BT, Frosch MP, Irizarry MC. Plasma A β levels do not reflect brain A β levels. *J Neuropathol Exp Neurol* 2007;66:264-71.
52. van Oijen M, Hofman A, Soares HD, Koudstaal PJ, Breteler MM. Plasma A β (1-40) and A β (1-42) and the risk of dementia: a prospective case-cohort study. *Lancet Neurol* 2006;5:655-60.
53. Pesaresi M, Lovati C, Bertora P, Mailland E, Galimberti D, Scarpini E, et al. Plasma levels of β -amyloid (1-42) in Alzheimer's disease and mild cognitive impairment. *Neurobiol Aging* 2006;27:904-5.
54. Graff-Radford NR, Crook JE, Lucas J, Boeve BF, Knopman DS, Ivnik RJ, et al. Association of low plasma A β 42/A β 40 ratios with increased imminent risk for mild cognitive impairment and Alzheimer disease. *Arch Neurol* 2007;64:354-62.
55. Blasko I, Jellinger K, Kemmler G, Krampla W, Jungwirth S, Wichart I, et al. Conversion from cognitive health to mild cognitive impairment and Alzheimer's disease: Prediction by plasma amyloid- β 42, medial temporal lobe atrophy and homocysteine. *Neurobiol Aging*. 2007 2008;29:1-11.
56. Oprisiu R, Serot JM, Godefroy O, Black SE, Fournier A. Plasma amyloid- β concentrations in Alzheimer's disease: an alternative hypothesis. *Lancet Neurol* 2006;5:1001-2.
57. Markesbery WR, Kryscio RJ, Lovell MA, Morrow JD. Lipid peroxidation is an early event in the brain in amnesic mild cognitive impairment. *Ann Neurol* 2005;58:730-5.
58. Montine TJ, Montine KS, McMahan W, Markesbery WR, Quinn JF, Morrow JD. F2-isoprostanes in Alzheimer and other neurodegenerative diseases. *Antioxid Redox Signal* 2005;7:269-75.
59. Padovani A, Borroni B, Colciaghi F, Pastorino L, Archetti S, Cottini E, et al. Platelet amyloid precursor protein forms in AD: a peripheral diagnostic tool and a pharmacological target. *Mech Ageing Dev* 2001;122:1997-2004.
60. Padovani A, Borroni B, Colciaghi F, Pettenati C, Cottini E, Agosti C, et al. Abnormalities in the pattern of platelet amyloid precursor protein forms in patients with mild cognitive impairment and Alzheimer disease. *Arch Neurol* 2002;59:71-5.
61. Borroni B, Colciaghi F, Caltagirone C, Rozzini L, Broglio L, Cattabeni F, et al. Platelet amyloid precursor protein abnormalities in mild cognitive impairment predict conversion to dementia of Alzheimer type: a 2-year follow-up study. *Arch Neurol* 2003;60:1740-4.
62. Davidsson P, Sjogren M. The use of proteomics in biomarker discovery in neurodegenerative diseases. *Dis Markers* 2005;21:81-92.
63. D'Ascenzo M, Relkin NR, Lee KH. Alzheimer's disease cerebrospinal fluid biomarker discovery: a proteomics approach. *Curr Opin Mol Ther* 2005;7:557-64.
64. Castano EM, Roher AE, Esh CL, Kokjohn TA, Beach T. Comparative proteomics of cerebrospinal fluid in neuropathologically-confirmed Alzheimer's disease and non-demented elderly subjects. *Neurol Res* 2006;28:155-63.
65. Davidsson P, Sjogren M. Proteome studies of CSF in AD patients. *Mech Ageing Dev* 2006;127:133-7.
66. Finehout EJ, Franck Z, Choe LH, Relkin N, Lee KH. Cerebrospinal fluid proteomic biomarkers for Alzheimer's disease. *Ann Neurol* 2007;61:120-9.
67. Simonsen AH, McGuire J, Podust VN, Davies H, Minthon L, Skoog I, et al. Identification of a novel panel of cerebrospinal fluid biomarkers for Alzheimer's disease. *Neurobiol Aging* 2007 (in press).
68. Simonsen AH, McGuire J, Hansson O, Zetterberg H, Podust VN, Davies HA, et al. Novel panel of cerebrospinal fluid biomarkers for the prediction of progression to Alzheimer dementia in patients with mild cognitive impairment. *Arch Neurol* 2007;64:366-70.
69. Hye A, Lynham S, Thambisetty M, Causevic M, Campbell J, Byers HL, et al. Proteome-based plasma biomarkers for Alzheimer's disease. *Brain* 2006;129:3042-50.
70. Ng TP, Chiam PC, Lee T, Chua HC, Lim L, Kua EH. Curry consumption and cognitive function in the elderly. *Am J Epidemiol* 2006;164:898-906.
71. Feng L, Ng TP, Chuah L, Niti M, Kua EH. Homocysteine, folate, and vitamin B-12 and cognitive performance in older Chinese adults: findings from the Singapore Longitudinal Ageing Study. *Am J Clin Nutr* 2006;84:1506-12.
72. Tham W, Auchus AP, Thong M, Goh ML, Chang HM, Wong MC, et al. Progression of cognitive impairment after stroke: one year results from a longitudinal study of Singaporean stroke patients. *J Neurol Sci* 2002;203:204:49-52.
73. Auchus AP, Chen CP, Sodagar SN, Thong M, Sng EC. Single stroke dementia: insights from 12 cases in Singapore. *J Neurol Sci* 2002;203:204:85-9.
74. Chong MS, Sahadevan S. Preclinical Alzheimer's disease: diagnosis and prediction of progression. *Lancet Neurol* 2005;4:576-9.
75. Chong MS, Lim WS, Sahadevan S. Biomarkers in preclinical Alzheimer's disease. *Curr Opin Investig Drugs* 2006;7:600-7.